Low ratio of S-adenosylmethionine to S-adenosylhomocysteine is associated with vitamin deficiency in Brazilian pregnant women and newborns

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ABSTRACT
Background: Pregnant women with low cobalamin concentrations are unable to provide the necessary amount of cobalamin to their fetuses. The effect of low maternal cobalamin concentrations on transmethylation metabolism in pregnant women and their newborns is unknown.
Objective: We investigated the relation between maternal and neonatal cobalamin concentrations and changes in total homocysteine (tHcy), S-adenosylmethionine (SAM), and S-adenosylhomocysteine (SAH).
Design: Hematologic data and concentrations of cobalamin, red blood cell folate, serum folate, tHcy, methylmalonic acid, SAM, SAH, and other metabolites were measured in 119 serum specimens from pregnant Brazilian women (gestational age: 37–42 wk) and their newborns’ placental veins at the time of delivery.
Results: The tHcy concentrations were higher in placental vein serum from newborns whose mothers had low cobalamin. Serum SAH concentrations were elevated and serum SAM and methionine concentrations were decreased in pregnant women with lower cobalamin concentrations. SAM:SAH was significantly decreased in both cobalamin-deficient pregnant women and their newborns.
Conclusions: Lower maternal cobalamin concentrations are associated with higher tHcy and lower SAM:SAH in newborns. Because SAM:SAH is closely linked with the activity of numerous enzymatic methylation reactions, these results suggest that methylation could be impaired in cobalamin-deficient pregnant women and their newborns.

KEY WORDS Cobalamin, folate, homocysteine, methylmalonic acid, S-adenosylhomocysteine, S-adenosylmethionine, pregnant women, newborns

INTRODUCTION
Cobalamin and folate are necessary for DNA synthesis and thus for cell division. Fetuses and pregnant women are in a state of rapid turnover that requires a high rate of DNA synthesis (1).
Methylcobalamin is a required coenzyme in the synthesis of methionine from homocysteine. This pathway involves the enzymatic transfer of a methyl group from 5-methyltetrahydrofolate to homocysteine by methionine synthase. In the absence of cobalamin, folate is “trapped” and cannot be recycled into the folate pool. Serum concentrations of total homocysteine (tHcy) are elevated in cobalamin and folate deficiency (2). It was theorized that impaired synthesis of methionine and therefore of S-adenosylmethionine (SAM), the most important methyl donor, plays a role in the pathophysiology of cobalamin deficiency. After donating a methyl group, SAM is converted to S-adenosylhomocysteine (SAH), which is hydrolyzed to homocysteine. Elevated concentrations of SAH can inhibit transmethylation reactions (3), which could also be deleterious in cobalamin deficiency. We showed previously that patients with severe cobalamin-deficient megaloblastic anemia have elevated serum SAH, which is corrected with cobalamin therapy (4).
Cobalamin is also a cofactor in the enzymatic conversion of l-methylmalonyl-coenzyme A to succinyl-coenzyme A. Impaired cobalamin status causes accumulation of serum methylmalonic acid (MMA; 2).
During pregnancy, the cell multiplication resulting from the enlargement of the uterus, placental development, and fetal growth increases cobalamin and folate requirements. It is well known that maternal serum cobalamin and folate concentrations decrease during pregnancy (5–8). Bruinse and van den Berg (6) reported that the cobalamin serum concentration fell gradually by ≈75–100 pmol/L during pregnancy. It is not known why cobalamin or folate decreases during gestation, but several factors, such as hemodilution, hormonal influences, and nutritional deficiency, could be responsible. The serum

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cortisol concentrations were an important variable for explaining the variance in cobalamin concentrations at the 34th wk of pregnancy (6). Cortisol, progesterone, and prolactin were associated with decreasing folate concentrations during pregnancy (6). Many studies showed relations between low folate concentrations, elevated tHcy, and pregnancy complications or adverse neonatal outcomes from neural tube defects (NTDs) and low birth weight (9–17).

Cobalamin deficiency was also related to the incidence of NTDs and birth complications. Thame et al (18) found an 11.8% preva lence of cobalamin deficiency (cobalamin <110.67 pmol/L) in Brazilian women with NTD-affected pregnancy. In addition, Adams et al (19) observed higher MMA concentrations in women with NTD-affected pregnancy than in women with unaffected pregnancy. However, the midtrimester MMA and tHcy values in their subjects were 25% and 34% lower than in female blood donors, which suggests that these metabolites, like the vitamin concentrations, decrease during normal gestation (19). A recent study showed that preterm birth was related to both hyperhomocysteinemia and cobalamin deficiency (20). The aim of this study was to investigate the relationship between maternal and neonatal cobalamin and folate status by analyzing the vitamin-dependent metabolites, including SAM and SAH, in pregnant women and in placental vein blood from their newborns.

**SUBJECTS AND METHODS**

**Subjects**

Blood was collected from 119 pregnant women at 2 public hospitals in Sorocaba City, Brazil, after admission for labor and up to 12 h before delivery. After expulsion of the placenta, the blood from placental veins was collected.

All deliveries occurred within the gestational age range of 37–42 wk. Women with a clinical diagnosis of metabolic diseases, renal insufficiency, increased serum liver enzymes, multiple gestations, and complications during delivery, including the birth of a premature newborn or a newborn with congenital malformation and anoxia, were excluded from the study.

Socioeconomic data (monthly per capita income, schooling, and occupation), obstetric status, and supplemental vitamin intake were assessed by questionnaire. Neonatal birth weight and Apgar score were collected. The protocol was approved by the local ethics committees (Brazil) and by the investigational review board at the University of Colorado, and written informed consent was obtained from all pregnant women before participation.

**Blood sampling**

Peripheral venous maternal blood and samples from placental veins were drawn into tubes with EDTA for measurements of the hemoglobin and whole blood folate concentration. Serum was obtained for measurements of concentrations of cobalamin, folate, tHcy, cystathionine, cysteine, methionine, SAM, SAH, MMA, 2-methylcitric acid, serine, glycine, N,N-dimethylglycine, and N-methylglycine. The clot tubes were cooled immediately at 4 °C and centrifuged 1 h after blood collection. The serum was frozen and sent to the United States frozen on dry ice and assayed within 1 y of collection.

**Biochemical measurements**

Red blood cell (RBC), leukocyte, and platelet counts; hemoglobin concentration; and hematocrit were measured on all samples with use of an electronic cell counter (Coulter STKS; Coulter Corp, Hialeah, FL). Serum folate concentrations were measured by using the ion capture method (IMx System; Abbott Laboratories, Abbott Park, IL). This method is based on binding by a specific folate-binding protein. RBC folate was measured with use of the same procedure after 1:20 dilution of a whole blood aliquot with lysis solution (Folate Lysis Reagent No. 9C13-60; Abbott Laboratories). RBC folate concentration was calculated as follows: folate content (in nmol/L)/hematocrit × dilution factor.

The serum cobalamin concentration was measured by 2 methods: method 1 used a totally automated kit (IMx System; Abbott Laboratories) with no boil pretreatment, and method 2 used a semiautomated kit (Immulite; Diagnostic Products Corp, Los Angeles), which involved sample pretreatment in a boil bath. Both methods use porcine intrinsic factor as a ligand.

Indexes of kidney and liver function were investigated with measurements of serum creatinine, aspartate aminotransferase, and alanine aminotransferase obtained by using kits (Dimension AR; Dade Behring, Newark, DE). Pregnant women with abnormalities in these indexes were excluded from our study.

The measurements of tHcy, MMA, and other amino acids were performed with the use of stable isotope dilution capillary gas chromatography–mass spectrometry (21–23). SAM and SAH were measured with use of a newly developed stable isotope dilution liquid chromatography–mass spectrometry method (24). The CV for intraassay precision (within-run) was 5% for SAM and 17% for SAH. The CV for interassay precision (run-to-run) was 7.4% for SAM and 22% for SAH (24).

**Statistical analysis**

All statistical analyses were carried out with use of SAS for WINDOWS software (version 6.12; SAS Institute Inc, Cary, NC), with the level of significance set at $P < 0.05$. The correlation between the concentrations of cobalamin measured by the 2 methods was assessed with use of the Pearson coefficient. The Bland-Altman analysis was used to study the agreement between methods of measuring cobalamin concentrations (25).

To establish a cobalamin deficiency cutoff in pregnant women and newborn groups, we used the highest quartile of maternal tHcy (>7.8 μmol/L), maternal MMA (>270 nmol/L), maternal SAH (>27 nmol/L), neonatal tHcy (>6.8 μmol/L), neonatal MMA (>384 nmol/L), and neonatal SAH (>67 nmol/L) and the lowest quartile of maternal (<2.6) and neonatal (<2.8) SAM: SAH. The maternal or neonatal variables were used alone or in combinations of variables as criteria of cobalamin deficiency in these groups.

The correlation between the concentrations of cobalamin and those of tHcy, MMA, and other metabolites in pregnant women and in newborns independently and the correlations of the concentrations between pregnant women and their respective newborns were determined by the Pearson coefficient. This coefficient was also used to measure the correlation between neonatal birth weight and maternal and neonatal biochemical measurements. The biochemical variables (ie, cobalamin, RBC folate, serum folate, tHcy, MMA, SAM, SAH, SAM:SAH, methionine,
cystathionine, 2-methylcitric acid, cysteine, glycine, serine, N,N-

dimethylglycine, and N-methylglycine), hematologic variables
(RBC count and hemoglobin concentration), neonatal birth
weight, and per capita income data were not consistent with a
normal distribution and were transformed in a logarithmic scale
for the statistical analysis. The maternal age data were not log
transformed.

The paired Student’s t test was used to compare the mean of
log-transformed data on the limits evaluated in pregnant women
and newborn groups. One-way analysis of variance was used to
compare the maternal and neonatal biochemical log-transformed
data from groups formed according to quartiles of maternal co-
balamin concentrations. When significant differences among
groups were observed, Tukey’s post hoc test was performed to
identify the significantly different group means.

Multiple linear and multiple logistic regression analysis
To assess the simultaneous relations between the various pre-
dictors of tHcy, MMA, and SAM:SAH in the pregnant women
and the newborns (as dependent variables), several models of
stepwise multiple linear regression analysis were used. For all
models, the biochemical variables were log transformed before
the analysis. The models were adjusted by covariates: maternal
age, racial group (whites compared with blacks or mixed groups),
and parity.

In model 1, the dependent variables were maternal tHcy,
MMA, and SAM:SAH. The independent variables included co-
balamin, RBC folate, serum folate, creatinine, SAM:SAH, and
tHcy for the dependent variables tHcy and SAM:SAH. The inde-
pendent variables for MMA were cobalamin, RBC folate, se-
rum folate, and creatinine. In model 2, the dependent variables
were neonatal tHcy, MMA, and SAM:SAH, and the independent
variables were the same maternal variables as in model 1. In
model 3, the dependent variables were the same as those in model
2, and the independent variables are the same as those in model
1 but were from the newborns. In model 4, the dependent vari-
ables were the same as those in models 2 and 3, and the inde-
pendent variables were those significant maternal and neonatal
variables in models 2 and 3, separately, for tHcy, MMA, and
SAM:SAH.

A multivariate logistic regression with stepwise analysis was
done with the same independent variables as were used in models
1–4 from the multiple linear regression analysis. The outcomes
were the following: maternal tHcy, >7.8 μmol/L; maternal
MMA, >270 nmol/L; maternal SAM:SAH, <2.6; neonatal
tHcy, >6.8 μmol/L; neonatal MMA, >384 nmol/L; and neo-

tal SAM:SAH, <2.8. These covariates of maternal age (year),
racial group (whites compared with blacks plus mixed racial
group), and parity were used in all of the multivariate logistic
regression analyses. The cutoff values for maternal and neonatal
outcomes were the highest quartiles for tHcy and MMA and the
lowest quartiles for SAM:SAH.

Receiver operator characteristic curve
The receiver operator characteristic (ROC) curve was used for
establishing the cobalamin cutoff indicators of metabolic cobal-
amin deficiency (26). The values of sensitivity and specificity for
each biochemical variable (tHcy, MMA, SAH, and SAM:SAH
or combinations of variables) were obtained by using contin-
guity tables that crossed the criteria and the number of subjects
below and above the cutoff value set in the ROC curves to
maximize both sensitivity and specificity. When ≥1 limit was
used as the criterion, it was considered altered if ≥1 of the limits
was altered. The area under the ROC curve tends to be near 1.0
for a sensitive and specific diagnostic method (26).

RESULTS
Population characteristics
The mean (±SD) age of the pregnant women was 25.5 ± 6.7
y (range: 15–44 y), gestational age was 39.4 ± 1.1 wk (range:
37–42 wk), and parity was 2.6 ± 1.7, including the current
pregnancy. The pregnant women were generally of low socio-

economic status without an occupation (74.8%) and without
basic schooling (42.4%). The mean (±SD) per capita income
was US$86.50 ± 65.20/mo. The cohort’s racial composition was
53.9% white (n = 64), 6.7% black (n = 8), 38.7% mixed black
and white (n = 46), and 0.8% Asian (n = 1).

Cobalamin assays
We measured the cobalamin concentrations by using 2 meth-

dods. Strong and significant correlations were found between the 2
methods in both the pregnant (r = 0.933, P < 0.001) and the new-
born (r = 0.963, P < 0.001) subjects (Figure 1). For the statistical
evaluations, we used concentrations measured by using method 1.

Simple correlations between vitamins and metabolites
Maternal biochemical values
The geometric means and 95% CIs of cobalamin, folate,
MMA, tHcy, and other metabolites of methionine metabolism
from maternal subjects are shown in Table 1. Significant relations
were found between cobalamin concentrations and tHcy (


P = 0.007, P = 0.001) in the newborns. There were weak but significant correlations were found between birth
cobalamin concentrations were low, and mean MMA and cystath-
ionine concentrations were relatively high in this population.
There were inverse correlations between maternal
cobalamin concentrations and tHcy (r = −0.186, P = 0.043)
SAM (r = 0.232, P = 0.013), and SAH (r = −0.248, P = 0.007)
concentrations and SAM:SAH (r = 0.293, P = 0.002). No sig-
nificant correlation was observed between maternal cobalamin
and MMA (r = −0.0106, P = 0.251). There were inverse corre-
lations between RBC folate and tHcy (r = −0.337, P < 0.001)
and between serum folate and tHcy (r = −0.485, P < 0.001).
No correlation was found between tHcy and cystathionine
concentrations in the pregnant subjects (r = 0.055, P = 0.553).
However, significant correlations were found between tHcy and
cysteine concentrations in the pregnant subjects (r = 0.245, P = 0.007).
No significant correlations were observed between birth
weight and maternal cobalamin, RBC folate, or serum folate
concentrations.

Placental vein biochemical values
The geometric means and 95% CIs of cobalamin, folate,
MMA, tHcy, and other biochemical variables from the placental
vein samples are shown in Table 1. Significant relations were
found between cobalamin concentrations and tHcy (r = −0.321,
P < 0.001) and MMA (r = −0.338, P < 0.001) in the newborns.
In contrast, no significant correlations were observed between
RBC folate and tHcy (r = −0.148, P = 0.123) or between serum
folate and tHcy (r = −0.154, P = 0.114).
There was no correlation between tHcy and cystathionine concentrations in the neonatal samples ($r = 0.056$, $P = 0.558$). However, a significant correlation was found between tHcy and cysteine concentrations ($r = 0.414$, $P < 0.001$). No significant correlations were observed between birth weight and neonatal cobalamin, RBC folate, or serum folate concentrations.

**Maternal and placental vein biochemical values**

SAM, SAH, MMA, methionine, cystathionine, 2-methylcitric acid, glycine, serine, $N,N$-dimethylglycine, and $N$-methylglycine concentrations were significantly higher in the placental samples. No significant difference was observed between maternal and neonatal SAM:SAH ($P = 0.605$). However, the neonatal tHcy and cysteine concentrations were significantly lower than were the maternal concentrations. Strong associations were observed between the maternal and neonatal variables for cobalamin, tHcy, MMA, cystathionine, $N,N$-dimethylglycine, and $N$-methylglycine (Table 1). Weaker relations were observed between maternal and neonatal SAM, maternal and neonatal SAH, and maternal and neonatal SAM:SAH (Table 1).

**Effect of maternal cobalamin concentrations on metabolites**

Data from pregnant women and placental vein samples were divided into quartiles according to maternal cobalamin concentrations: quartile 1 (group 1), cobalamin $\leq 102$ pmol/L ($n = 29$); quartile 2 (group 2), cobalamin $\leq 128$ pmol/L ($n = 29$); quartile 3 (group 3), cobalamin $\leq 162$ pmol/L ($n = 30$); and quartile 4 (group 4), cobalamin $> 162$ pmol/L ($n = 30$), as shown in Table 2.

Maternal age was not significantly different across different cobalamin quartiles. The pregnant women from the lowest quartile (group 1) had lower mean RBC counts ($3.82 \times 10^9$/L) and hemoglobin concentration (109.7 g/L) than did pregnant women in the highest quartile (group 4) ($4.15 \times 10^9$/L and 119.1 g/L, respectively). However, no significant difference was found in RBC counts or hemoglobin concentrations between the respective newborn groups.

Serum folate but not RBC folate concentration was lower in women from group 1 than in women from group 4. The tHcy concentrations did not increase significantly in women with the lowest cobalamin values ($P = 0.112$). The placental vein cobalamin
TABLE 1
Geometric means, 95% CIs, and Pearson correlations between maternal and neonatal placental vein data

<table>
<thead>
<tr>
<th>Variables</th>
<th>Pregnant subjects</th>
<th>Neonatal placental vein</th>
<th>Pearson correlation coefficient ($r$)</th>
<th>$p$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Cobalamin, pmol/L ($n = 117$)</td>
<td>130 (122, 138)</td>
<td>205 (186, 225)</td>
<td>0.570</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Red blood cell folate, nmol/L ($n = 116$)</td>
<td>643 (591, 701)</td>
<td>1108 (1033, 1188)</td>
<td>0.488</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Serum folate, nmol/L ($n = 112$)</td>
<td>12.9 (12.0, 14.0)</td>
<td>30.9 (29.8, 32.1)</td>
<td>0.409</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Total homocysteine, μmol/L ($n = 110$)</td>
<td>6.5 (6.1, 6.9)</td>
<td>5.8 (5.4, 6.1)</td>
<td>0.753</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>MMA, nmol/L ($n = 110$)</td>
<td>200 (185, 216)</td>
<td>308 (289, 328)</td>
<td>0.693</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>SAM, nmol/L ($n = 105$)</td>
<td>75 (70, 81)</td>
<td>176 (164, 190)</td>
<td>0.219</td>
<td>0.024</td>
</tr>
<tr>
<td>SAH, nmol/L ($n = 107$)</td>
<td>22 (20, 23)</td>
<td>52 (48, 57)</td>
<td>0.219</td>
<td>0.023</td>
</tr>
<tr>
<td>Ratio of SAM to SAH ($n = 105$)</td>
<td>3.5 (3.1, 3.9)</td>
<td>3.4 (3.1, 3.7)</td>
<td>0.288</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Methionine, μmol/L ($n = 110$)</td>
<td>16.9 (16.2, 17.5)</td>
<td>26.3 (25.4, 27.3)</td>
<td>0.394</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Cystathionine, nmol/L ($n = 110$)</td>
<td>197 (183, 212)</td>
<td>314 (290, 340)</td>
<td>0.728</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>2-Methylcitric acid, nmol/L ($n = 109$)</td>
<td>115 (109, 121)</td>
<td>177 (170, 185)</td>
<td>0.478</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Cysteine, μmol/L ($n = 110$)</td>
<td>191 (185, 196)</td>
<td>181 (176, 187)</td>
<td>0.432</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Glycine, μmol/L ($n = 110$)</td>
<td>190 (182, 199)</td>
<td>281 (271, 292)</td>
<td>0.474</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>Serine, μmol/L ($n = 110$)</td>
<td>136 (131, 141)</td>
<td>155 (151, 160)</td>
<td>0.417</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>N,N-Dimethylglycine, μmol/L ($n = 110$)</td>
<td>2.8 (2.5, 3.1)</td>
<td>3.7 (3.4, 4.1)</td>
<td>0.936</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>N-methylglycine, μmol/L ($n = 110$)</td>
<td>0.6 (0.6, 0.7)</td>
<td>0.9 (0.9, 1.0)</td>
<td>0.826</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

1 All values are geometric $\bar{x}$; 95% CIs in parentheses. MMA, methylenalonic acid; SAM, S-adenosylmethionine; SAH, S-adenosylhomocysteine.
2 $n = 113–119$ for the different variables.
3 $n = 111–117$ for the different variables.
4 Calculated on the log-transformed variables.

Concentrations from maternal groups 1, 2, and 3 were significantly lower than those from maternal group 4 (highest cobalamin quartile; Table 2), whereas there were no changes in folate concentrations between groups. Neonatal tHcy concentrations were higher in the groups with lower maternal cobalamin. Serum SAM and methionine concentrations were decreased, and serum SAH concentrations were elevated in pregnant women with lower cobalamin concentrations. SAM:SAH was significantly decreased in both pregnant subjects and placental vein samples in the lowest group. Maternal and neonatal cysteine concentrations were not different according to the quartile of maternal cobalamin.

TABLE 2
Maternal and placental vein biochemical variables divided by maternal cobalamin quartile

<table>
<thead>
<tr>
<th>Variables</th>
<th>Quartile of maternal serum cobalamin 1, 2, 3, 4</th>
<th>$p$</th>
</tr>
</thead>
<tbody>
<tr>
<td>Maternal cobalamin (pmol/L)</td>
<td>84 (79, 90)</td>
<td></td>
</tr>
<tr>
<td>Neutonal cobalamin (pmol/L)</td>
<td>149 (127, 174) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal RBC folate (nmol/L)</td>
<td>614 (505, 745) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal RBC folate (nmol/L)</td>
<td>995 (874, 1133) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal serum folate (nmol/L)</td>
<td>10.7 (9.2, 12.4) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal serum folate (nmol/L)</td>
<td>30.3 (27.8, 33.2) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal Hcy (μmol/L)</td>
<td>7.1 (6.0, 8.3) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal Hcy (μmol/L)</td>
<td>7.0 (6.0, 8.1) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal MMA (nmol/L)</td>
<td>199 (169, 233) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal MMA (nmol/L)</td>
<td>319 (284, 359) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal SAM (nmol/L)</td>
<td>63 (55, 73) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal SAM (nmol/L)</td>
<td>161 (131, 199) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal SAH (nmol/L)</td>
<td>26 (22, 30) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal SAH (nmol/L)</td>
<td>59 (47, 74) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal SAM/SAH</td>
<td>2.5 (2.0, 3.1) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal SAM/SAH</td>
<td>2.7 (2.4, 3.1) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal methionine (μmol/L)</td>
<td>15.5 (14.2, 16.8) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal methionine (μmol/L)</td>
<td>27.1 (25.1, 29.4) $^a$</td>
<td></td>
</tr>
<tr>
<td>Maternal cysteine (μmol/L)</td>
<td>181 (171, 192) $^a$</td>
<td></td>
</tr>
<tr>
<td>Neutonal cysteine (μmol/L)</td>
<td>186 (173, 199) $^a$</td>
<td></td>
</tr>
</tbody>
</table>

1 All values are geometric $\bar{x}$; 95% CI in parentheses. RBC, red blood cell; tHcy, total homocysteine; MMA, methylenalonic acid; SAM, S-adenosylmethionine; SAH, S-adenosylhomocysteine. Values in a row with different superscript letters are significantly different, $P < 0.05$ (Tukey’s test).
2 Quartile 1, ≤102 pmol/L; quartile 2, 103–128 pmol/L; quartile 3, 129–162 pmol/L; quartile 4, ≥163 pmol/L.
3 ANOVA performed on the log-transformed variables.
4 Statistical analysis was not evaluated for this variable.
Predictors of metabolite variables

The association between various maternal and neonatal biochemical variables and the outcomes for tHcy, MMA, and SAM:SAH in mothers and newborns by stepwise multiple linear regression analysis is shown in Table 3. The odds ratio for elevated tHcy and MMA or low SAM:SAH in mothers and neonates by stepwise multiple logistic regression analysis is shown in Table 4. Two categories for independent variables were used in multivariate logistic regression analysis.

In both linear and logistic regression analyses, we tested maternal determinants of maternal MMA, tHcy, and SAM:SAH (model 1); maternal determinants of neonatal MMA, tHcy, and SAM:SAH (model 2); neonatal determinants of neonatal MMA, tHcy, and SAM:SAH (model 3); and the combined neonatal and maternal determinants of neonatal MMA, tHcy, and SAM:SAH (model 4). Maternal serum folate concentration was an independent inverse predictor of maternal tHcy concentrations. The partial $R^2$ for this regression indicated that maternal serum folate concentrations explained 20.7% of the variance in maternal tHcy concentrations (Table 3). Pregnant women with low maternal serum folate ($<9.3 \text{ nmol/L}$) had a 5.8-fold greater risk of higher tHcy concentrations than did pregnant women with higher serum folate concentrations (Table 4). The maternal MMA concentrations were predicted only by maternal serum creatinine (partial $R^2 = 0.0740$; Table 3). The odds ratio for elevated maternal MMA for pregnant women with serum creatinine $\geq 0.7 \text{ mg/dL}$ was 4.8 (95% CI: 1.3, 17.8; Table 4). Maternal SAM:SAH was predicted by maternal cobalamin concentration (Table 3). The pregnant women with lower cobalamin concentrations had a 7.2-fold greater risk of lower maternal SAM:SAH than did pregnant women with higher serum cobalamin concentrations (Table 4).

The maternal cobalamin and maternal serum folate concentrations inversely predicted the neonatal tHcy concentrations (model 2). The partial $R^2$ values for these regressions indicate that maternal cobalamin accounts for 20.5% of the variance in neonatal tHcy concentrations, and maternal serum folate accounts for only 5.5% of the variance in neonatal tHcy concentrations. In model 3, which tested only neonatal independent variables, the neonatal cobalamin concentration and SAM:SAH were inversely associated with neonatal tHcy concentrations. In contrast to the maternal cobalamin, the neonatal cobalamin and neonatal SAM:SAH explained less of the variance in neonatal tHcy. Model 4 explores the significant maternal and neonatal independent variables from models 2 and 3. Model 4 showed that there was no increase in the summed value of partial $R^2$ when model 2 was compared with model 4 (26.0% and 24%, respectively). The same determinant variables from model 2 were selected (maternal cobalamin and maternal serum folate), and they were inversely associated with neonatal tHcy concentrations in model 4 (Table 3).

The predictors of neonatal MMA concentrations were maternal cobalamin and maternal serum creatinine in model 2, neonatal cobalamin in model 3, and neonatal cobalamin in model 4. The

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**Table 3**

Association between maternal and neonatal biochemical variables and total homocysteine (tHcy), methylmalonic acid (MMA), and the ratio of S-adenosylmethionine (SAM) to S-adenosylhomocysteine (SAH) in mothers and children by stepwise multiple linear regression analysis.

<table>
<thead>
<tr>
<th>Model and dependent variable</th>
<th>Step</th>
<th>Independent variables</th>
<th>$\beta$</th>
<th>Partial $R^2$</th>
<th>$P$</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Maternal tHcy</td>
<td>Maternal SF</td>
<td>-0.373</td>
<td>0.207</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>Maternal MMA</td>
<td>Maternal SC</td>
<td>0.609</td>
<td>0.074</td>
<td>0.003</td>
</tr>
<tr>
<td></td>
<td>Maternal SAM:SAH</td>
<td>Maternal cobalamin</td>
<td>0.504</td>
<td>0.089</td>
<td>0.001</td>
</tr>
<tr>
<td>2</td>
<td>Neonatal tHcy</td>
<td>Maternal cobalamin</td>
<td>-0.357</td>
<td>0.205</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>Maternal MMA</td>
<td>Maternal cobalamin</td>
<td>-0.211</td>
<td>0.055</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>Neonatal SAM:SAH</td>
<td>Maternal cobalamin</td>
<td>0.353</td>
<td>0.044</td>
<td>0.026</td>
</tr>
<tr>
<td>3</td>
<td>Neonatal tHcy</td>
<td>Neonatal cobalamin</td>
<td>-0.195</td>
<td>0.094</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>Neonatal MMA</td>
<td>Neonatal cobalamin</td>
<td>-0.128</td>
<td>0.038</td>
<td>0.033</td>
</tr>
<tr>
<td></td>
<td>Neonatal SAM:SAH</td>
<td>Neonatal cobalamin</td>
<td>-0.227</td>
<td>0.129</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td>4</td>
<td>Neonatal tHcy</td>
<td>Neonatal cobalamin</td>
<td>-0.345</td>
<td>0.194</td>
<td>&lt;0.001</td>
</tr>
<tr>
<td></td>
<td>Neonatal MMA</td>
<td>Neonatal cobalamin</td>
<td>-0.194</td>
<td>0.046</td>
<td>0.012</td>
</tr>
<tr>
<td></td>
<td>Neonatal SAM:SAH</td>
<td>Neonatal cobalamin</td>
<td>-0.225</td>
<td>0.132</td>
<td>&lt;0.001</td>
</tr>
</tbody>
</table>

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1 All variables were log transformed. The regression models were adjusted by maternal age, racial group, and parity. The regression parameter estimate ($\beta$), partial $R^2$, and $P$ values are for the independent variables within each model. SF, serum folate; SC, serum creatinine.

2 Model 1: Maternal variables are used as determinants of maternal tHcy, MMA, and SAM:SAH. Independent variables for tHcy and SAM:SAH include maternal RBC and serum concentrations of folate, cobalamin, creatinine, tHcy, and SAM:SAH. For MMA, the independent variables are maternal RBC and serum concentrations of folate, cobalamin, and creatinine. Model 2: Maternal variables are used as determinants of neonatal tHcy, MMA, and SAM:SAH. Independent variables are the same as in model 1. Model 3: Neonatal variables are used as determinants of neonatal tHcy, MMA, and SAM:SAH. Independent variables are the same as in model 1, but from the newborn. Model 4: Neonatal and maternal variables that were significant in model 2 or model 3 are used as determinants of neonatal tHcy, MMA, and SAM:SAH.
maternal cobalamin concentrations explained 3.5% of the variance in neonatal SAM:SAH in model 2 (Table 3), and pregnant women with lower cobalamin concentrations (<102 pmol/L) had a 5.0-fold greater risk of lower neonatal SAM:SAH than did pregnant women with higher serum cobalamin concentrations (Table 4). The neonatal serum folate and neonatal tHcy concentrations were inversely associated with neonatal SAM:SAH in models 3 and 4, and these variables explained 7.6% and 5.7% of the variance, respectively, in neonatal SAM:SAH (Table 3).

There were weak but significant inverse correlations between SAM:SAH and serum folate (r = −0.272, P = 0.005) and tHcy (r = −0.226, P = 0.017) in the newborns. The higher value of the regression parameter estimate (β = −0.853) for serum folate in models 3 and 4 of SAM:SAH as dependent variable could be explained by values in a particular range of folate distribution. Newborns with higher tHcy concentrations (>6.8 μmol/L) had a 3.2-fold greater risk of lower SAM:SAH than did newborns with lower tHcy concentrations, and the pregnant women with lower cobalamin concentrations had a 5.0-fold greater risk of delivery of a newborn with a lower SAM:SAH than did those pregnant women with higher cobalamin concentrations (Table 4).

**Establishment of cobalamin deficiency cutoffs in pregnant women and newborn groups**

We used ROC curves to establish a cobalamin deficiency cutoff in pregnant women and newborn groups (26). In pregnant women, the combination of tHcy concentrations and SAM:SAH was the best criteria (as shown by a combination of higher sensitivity and specificity) for detection of cobalamin deficiency (Table 5). When used as a single factor, tHcy had the best combination of sensitivity and specificity and showed the same cobalamin cutoff (<132 pmol/L) as was obtained in the combination of tHcy and SAM:SAH. According to this cutoff, 63 (52.9%) pregnant women in our study had cobalamin deficiency.

No significant differences were found among the values of area under ROC curves for all criteria used in the newborn group. The combination of tHcy and MMA concentrations showed the highest value of area under ROC curve (0.675; 95% CI: 0.567, 0.782) and a good sensitivity (69.0%) and specificity (62.7%). When used as single limits, tHcy and MMA also showed a good sensitivity and specificity. However, the cobalamin cutoff for MMA alone as the limit was lower than the cutoff values for tHcy alone and the combination of tHcy and MMA concentrations. By using the cobalamin cutoff (<194 pmol/L) for newborns, we found 54 (49.5%) newborns in our study who had cobalamin deficiency.

**DISCUSSION**

We studied the cobalamin and folate status of a group of socioeconomically disadvantaged Brazilian women in labor who likely were at risk of vitamin deficiency because of infrequent vitamin supplementation and the low animal protein content of their daily diet. We found that the mean maternal serum cobalamin concentration was much lower than values reported from the United States and Europe (6–8, 19, 27–30) and similar to a concentration detected in our study was much lower than values reported from Ireland (31). Most of the serum cobalamin concentrations detected in our study were <180 pmol/L, the value cited by Bruinse and van den Berg (6) to indicate marginal or deficient vitamin stores. The lower cobalamin concentrations were found by using 2 separate methods of quantitation, which correlated...
Newborns different superscript letters are significantly different, SAH is higher, and SAM:SAH is lower in women in the lowest study was probably adequate, unlike their cobalamin status. We conclude that the folate status of the Brazilian mothers in the current study was similar to values obtained in previous French (29), Spanish (30), Canadian (37), and US (38) investigations. Studies from Ireland (31) and the Netherlands (39) found much higher mean tHcy and lower folate values than we found. Therefore, we conclude that the folate status of the Brazilian mothers in the current study was probably adequate, unlike their cobalamin status.

For the first time, we showed that maternal SAM is lower, SAH is higher, and SAM:SAH is lower in women in the lowest cobalamin quartile than in those in the other quartiles. Because these subjects also had lower serum folate, it is possible that a combined deficiency exacerbated the abnormalities, although the multiple linear regression model showed that only the maternal cobalamin was a predictor of the maternal SAM:SAH. Because we previously showed that SAH is increased and SAM:SAH is decreased in Brazilian patients with severe cobalamin-deficient megaloblastic anemia (4), we interpret these changes in the pregnant subjects as showing biochemically significant cobalamin deficiency. This maternal cobalamin deficiency apparently causes neonatal cobalamin deficiency, because the neonatal cobalamin values were strongly dependent on the maternal values. The tHcy, MMA, and SAM:SAH were also abnormal in the newborns of mothers in the lowest cobalamin quartile. The inverse relations between neonatal tHcy, MMA, cobalamin, and serum folate concentrations and SAM:SAH and the maternal cobalamin and serum folate concentrations confirm that there were metabolic consequences of deficiency for the newborns. Cobalamin was shown to be an important determinant of neonatal tHcy in other studies (28, 31).

It appears that the analysis of SAM, SAH, and SAM:SAH might be of diagnostic utility in assessing vitamin deficiency in pregnancy. We were very careful to prepare the samples in a uniform manner and without allowing room-temperature storage or incubation at any time from phlebotomy to analysis. This care is important because we showed that SAH increases and SAM:SAH decreases after relatively short intervals of room-temperature or 4 °C incubation of samples (24). Further investigations of SAM and SAH after stringent sample preparation and storage conditions should be pursued in other populations with poor intake of cobalamin and folate.

TABLE 5
Cobalamin cutoffs for pregnant women and newborns obtained by using the total homocysteine (tHcy), methylmalonic acid (MMA), and S-adenosylhomocysteine (SAH) concentrations and the ratio of S-adenosylmethionine (SAM) to SAH as indicators of cobalamin deficiency.

<table>
<thead>
<tr>
<th>Criteria</th>
<th>Area under ROC curve</th>
<th>Sensitivity, specificity</th>
<th>Cobalamin cutoffs</th>
</tr>
</thead>
<tbody>
<tr>
<td>Pregnant subjects</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>tHcy</td>
<td>0.598 (0.483, 0.714)a,b</td>
<td>75.0, 54.4</td>
<td>132</td>
</tr>
<tr>
<td>MMA</td>
<td>0.497 (0.380, 0.614)a</td>
<td>48.3, 66.3</td>
<td>116</td>
</tr>
<tr>
<td>SAH</td>
<td>0.597 (0.472, 0.722)b</td>
<td>78.6, 42.5</td>
<td>141</td>
</tr>
<tr>
<td>SAM:SAH</td>
<td>0.724 (0.613, 0.836)b</td>
<td>78.6, 40.0</td>
<td>128</td>
</tr>
<tr>
<td>tHcy and MMA</td>
<td>0.544 (0.439, 0.650)b</td>
<td>64.6, 54.3</td>
<td>133</td>
</tr>
<tr>
<td>tHcy and SAM:SAH</td>
<td>0.682 (0.583, 0.782)b</td>
<td>75.0, 62.9</td>
<td>132</td>
</tr>
<tr>
<td>tHcy, MMA, and SAM:SAH</td>
<td>0.659 (0.559, 0.760)b</td>
<td>66.8, 64.8</td>
<td>133</td>
</tr>
<tr>
<td>tHcy, SAM, and SAM:SAH</td>
<td>0.661 (0.561, 0.761)b</td>
<td>72.4, 60.0</td>
<td>135</td>
</tr>
<tr>
<td>tHcy, MMA, SAH, and SAM:SAH</td>
<td>0.649 (0.545, 0.755)b</td>
<td>66.7, 65.2</td>
<td>134</td>
</tr>
<tr>
<td>Newborns</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>tHcy</td>
<td>0.645 (0.522, 0.768)a</td>
<td>72.0, 59.5</td>
<td>191</td>
</tr>
<tr>
<td>MMA</td>
<td>0.649 (0.523, 0.774)a</td>
<td>61.5, 71.1</td>
<td>164</td>
</tr>
<tr>
<td>SAH</td>
<td>0.528 (0.397, 0.659)a</td>
<td>64.0, 53.6</td>
<td>195</td>
</tr>
<tr>
<td>SAM:SAH</td>
<td>0.496 (0.360, 0.632)a</td>
<td>48.0, 64.3</td>
<td>167</td>
</tr>
<tr>
<td>tHcy and MMA</td>
<td>0.675 (0.567, 0.782)a</td>
<td>69.0, 62.7</td>
<td>194</td>
</tr>
<tr>
<td>tHcy and SAM:SAH</td>
<td>0.580 (0.464, 0.696)a</td>
<td>62.5, 60.9</td>
<td>191</td>
</tr>
<tr>
<td>tHcy, MMA, and SAM:SAH</td>
<td>0.614 (0.506, 0.722)a</td>
<td>52.8, 75.0</td>
<td>167</td>
</tr>
<tr>
<td>tHcy, SAM, and SAM:SAH</td>
<td>0.600 (0.490, 0.710)a</td>
<td>67.4, 61.9</td>
<td>195</td>
</tr>
<tr>
<td>tHcy, MMA, SAH, and SAM:SAH</td>
<td>0.622 (0.516, 0.729)a</td>
<td>64.3, 64.2</td>
<td>195</td>
</tr>
</tbody>
</table>

1 ROC, receiver operator characteristic. When ≥ 1 limit was used in the criteria, it was considered altered if ≥ 1 of the limits was altered. Values with different superscript letters are significantly different, P < 0.05 (ROC curves comparison test).
2 β, 95% CI in parentheses (all such values).
In this study, we drew blood from placental veins, which is a noninvasive method of obtaining blood from the neonatal circulation. The values obtained in this investigation were very similar to those previously obtained from studies of cord blood (40). The elevated serum SAH and low SAM:SAH found in the newborns and the mothers with the lowest cobalamin values could reflect an abnormal methylation status in the tissues (41). The mothers in the lowest quartile of cobalamin values were more severely anemic than were the rest of the mothers, but there was no apparent effect of vitamin status on the neonatal weight or other limits. Future studies should be directed toward following the development and cognitive abilities of these children. Children raised on macrobiotic diets, although admittedly much more severely cobalamin deficient than the subjects in this study, have been shown to have growth and cognitive impairment (42). DNA hypomethylation associated with elevated SAH was reported in vascular disease (43, 44). Such studies should be pursued in vitamin-deficient mother-newborn pairs.

The serum SAM:SAH is greatly dependent on the renal status of the person (24, 45). We previously showed that the urinary fractional excretion of SAM is close to 100% and SAH is of the person (24, 45). We previously showed that the urinary vitamin-deficient mother-newborn pairs. The serum SAM:SAH has been shown to have growth and cognitive impairment (42). DNA hypomethylation associated with elevated SAH was reported in vascular disease (43, 44). Such studies should be pursued in vitamin-deficient mother-newborn pairs.

We studied women attending public hospitals who had poor socioeconomic status and who likely consumed diets low in animal protein, which is the dietary source of cobalamin. Because this vitamin is not present in the plant protein sources (beans and grains) commonly eaten by this cohort, its members are likely cobalamin deficient on a dietary basis, although we did not access intake. Parasitic or bacterial infections could also play a role in cobalamin deficiency in this population. In addition, the pregnant women with low cobalamin concentrations had low serum folate concentrations, also probably because of low intake. However, lower folate concentrations could also result from the fact that cobalamin deficiency can interfere with folate metabolism. In the absence of cobalamin, methylfolate is trapped and cannot be recycled into the folate pool.

Cobalamin deficiency might prove to be an important public health problem in areas of the world where animal protein foods are not widely consumed for reasons of expense and scarcity, because lower serum cobalamin concentrations in pregnant women seem to have metabolic effects on their newborns at birth. Studies should be performed to show whether routine supplementation of vitamins that include both cobalamin and folic acid would be beneficial in such populations in improving maternal and child health.

We thank Helymar C Machado for statistical analysis. EMG-S was the coordinator of this study in Brazil and was responsible for all of its phases (planning, collecting of the samples, interviews, analysis of the data, interpretation of the values, and preparation of the manuscript). OM, SP, and RAP conducted the collecting of blood samples (from pregnant women and from placenta veins), interviewed the pregnant women, and processed the blood-samples right after blood collection. LFSN was the obstetrician who participated in planning and executing this study in the Hospital Regional and in Santa Lucinda Hospital. VDA participated in the preparation of the manuscript. SPI supervised and coordinated sample analysis. SP and OEM measured the concentrations of cobalamin, folate, ferritin, and hemoglobin at the hematology laboratory of the Clinical and Toxicology Analyses Department of the University of São Paulo. The Brazilian authors have no conflicts of interest. SPS and RHA were responsible for the assays of the amino acids, SAM, and SAH in their laboratories and also provided interpretation of the values and participated in the preparation of the manuscript. The University of Colorado and SPS and RHA hold patents on various aspects of the use of homocysteine and methylmalonic acid in the diagnosis of cobalamin or folate deficiency. A company was formed at the University of Colorado to perform the assays.

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